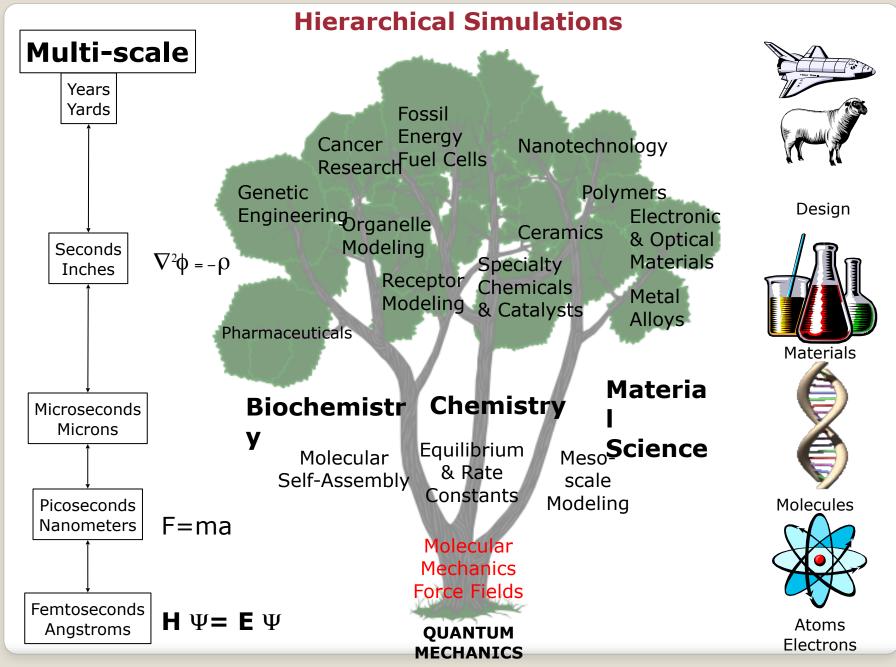
Biomolecular simulations

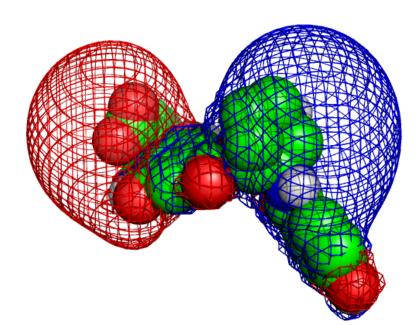
Patrice Koehl



© W.A. Goddard III, M. Blanco,

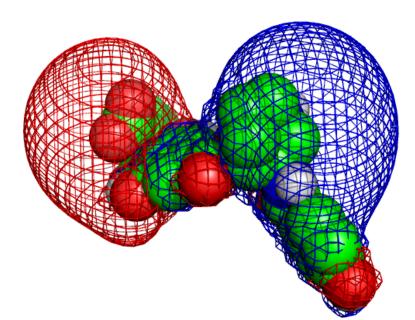
Biomolecular Simulations

- Molecular Mechanics force fields
- Energy Minimization
- Molecular dynamics
- Monte Carlo methods



Biomolecular Simulations

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The two major assumptions in molecular simulations

1. Born-Oppenheimer approximation

"the dynamics of electrons is so fast that they can be considered to react instantaneously to the motion of their nuclei"

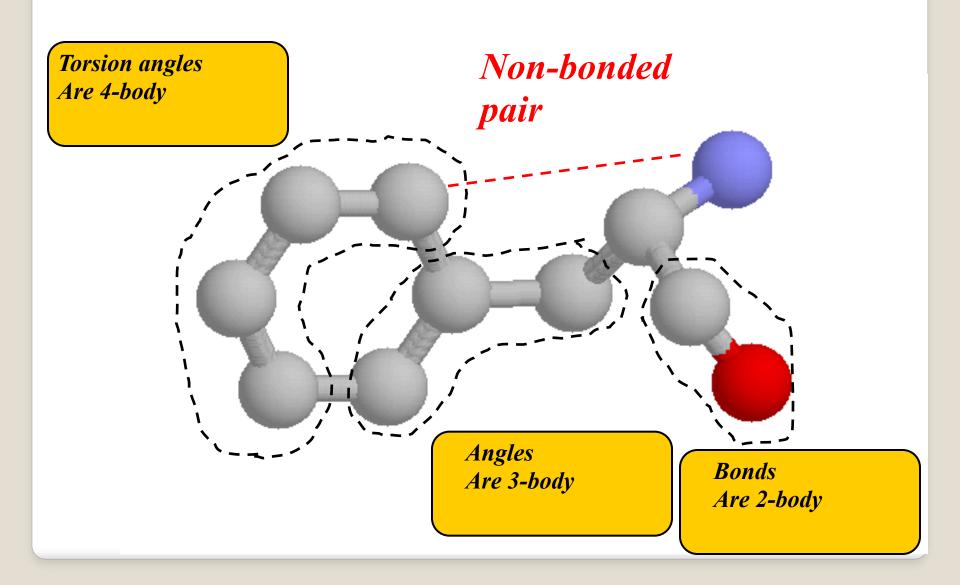
2. Classical mechanics

"The nuclei are treated as point particles that follow the classical laws of mechanics."

What is an atom?

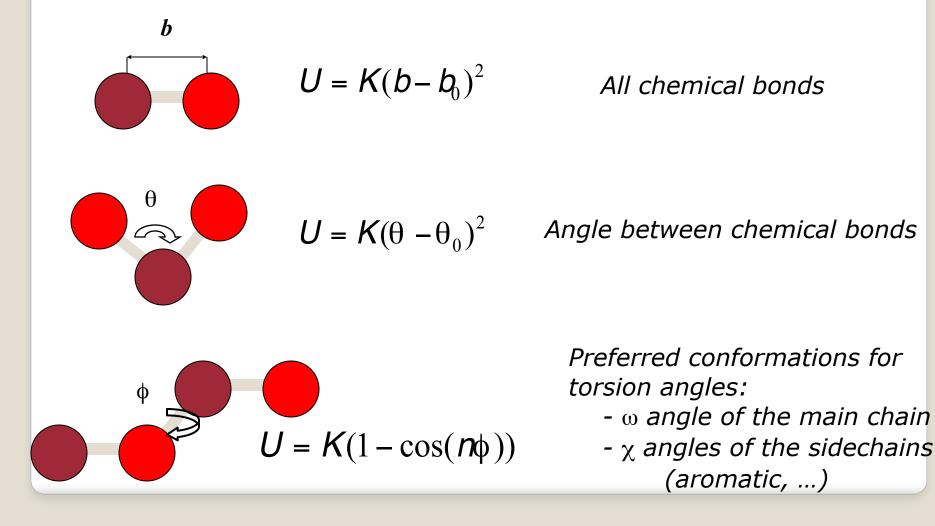
- Classical mechanics: a point particle
- Defined by its position (x,y,z) and its mass
- May carry an electric charge (positive or negative), usually partial (less than an electron)

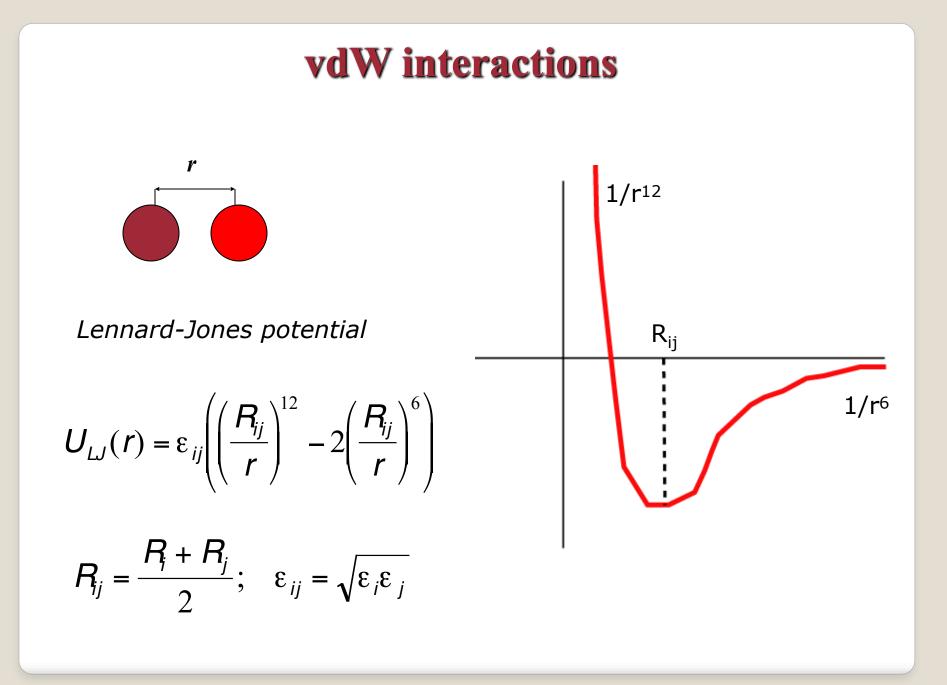
Atomic interactions



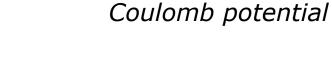
Atomic interactions

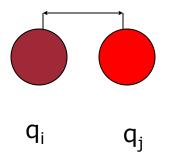
Strong valence energies





Electrostatics interactions

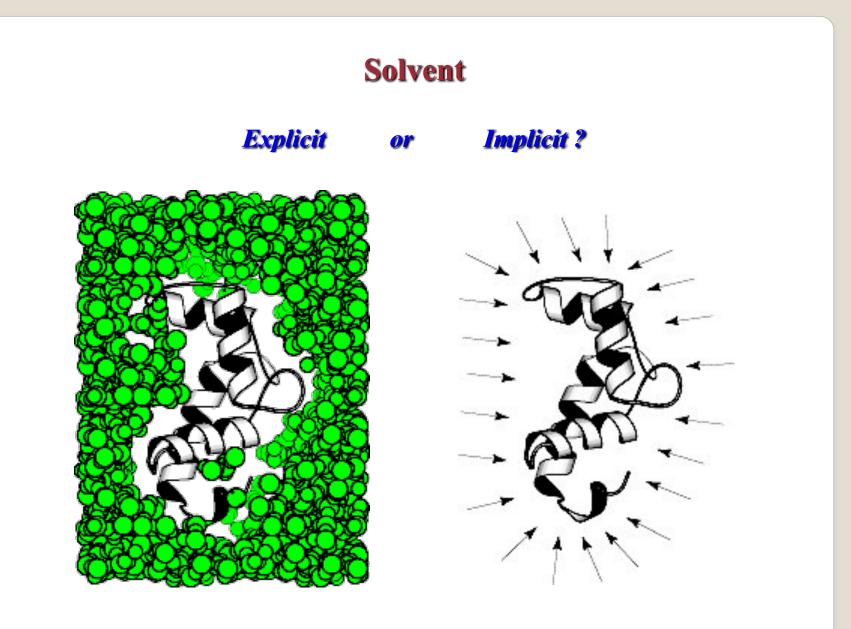




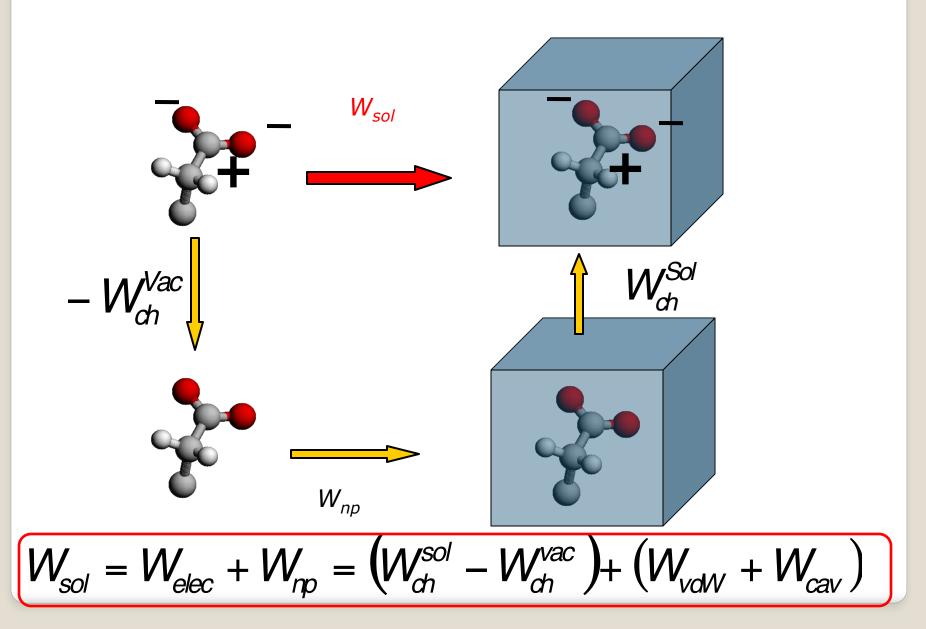
r

 $U(r) = \frac{1}{4\pi\varepsilon_0\varepsilon} \frac{q_i q_j}{r}$

Computing energy $=\sum_{all\ bonds}\frac{1}{2}K_{b}(b-b_{0})^{2}$ **Torsion angles** + $\sum_{\text{all angles}} \frac{1}{2} K_{\theta} (\theta - \theta_{0})^{2}$ Non-bonded Are 4-body pair + $\sum K_{\phi} [1 - \cos(n\phi)]$ all forsions + $\sum_{i, j \text{ nonbonded}} \varepsilon_{ij} \left[\left(\frac{R_{ij}}{r_{ij}} \right)^{12} - 2 \left(\frac{R_{ij}}{r_{ij}} \right)^{6} \right]$ Angles **Bonds** Are 3-body Are 2-body + $\sum_{i, j \text{ nonbonded}} \frac{q_i q_j}{4\pi \varepsilon_0 \varepsilon r_{ii}}$



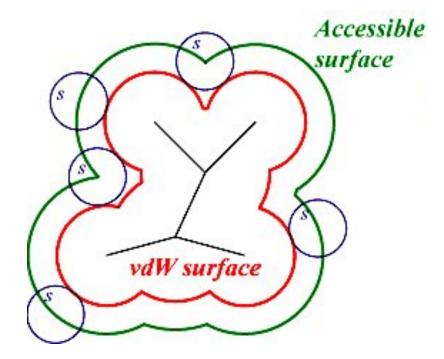
Solvation Free Energy



The SA model

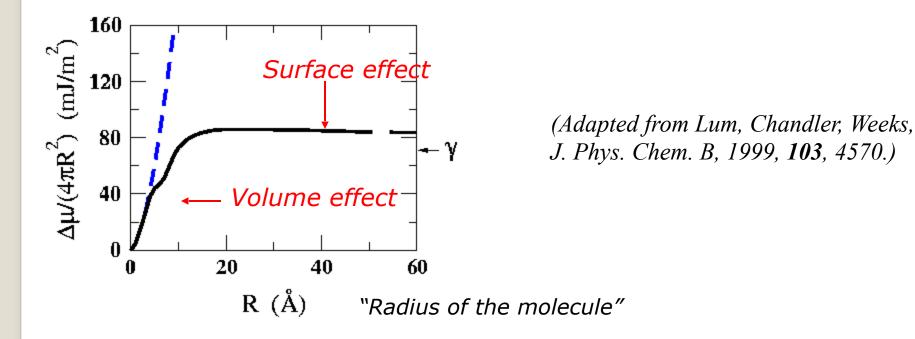
Surface area potential

$$W_{np} = W_{cav} + W_{vdW}$$
$$= \sum_{k=1}^{N} \sigma_k SA_k$$



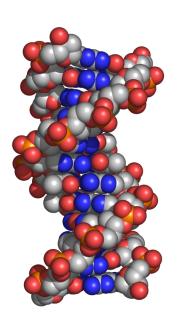
Eisenberg and McLachlan, (1986) Nature, 319, 199-203

Hydrophobic potential: Surface Area, or Volume?

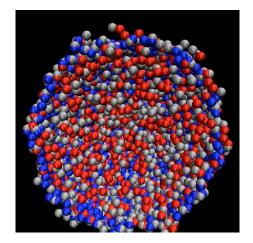


For proteins and other large bio-molecules, use surface

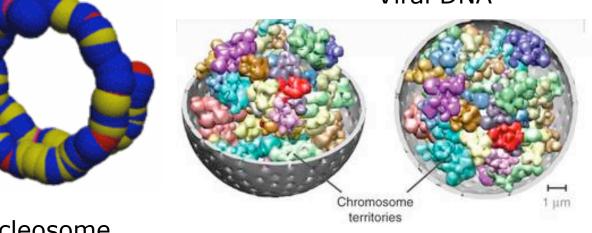
Sphere Representations in Biology







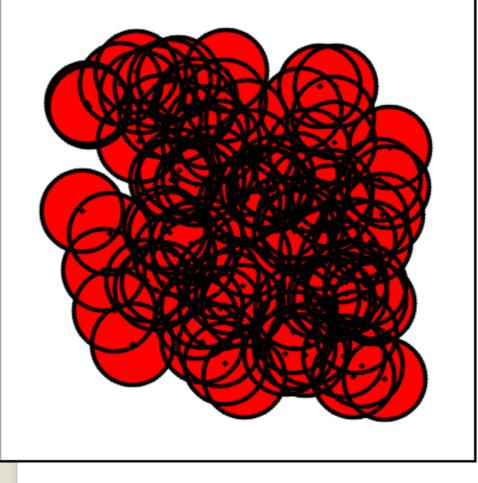
Viral DNA

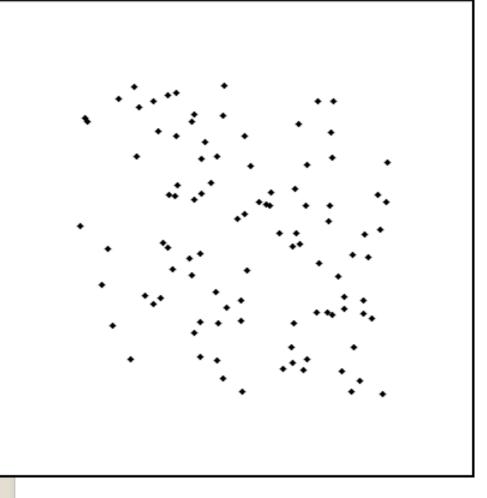


DNA

Nucleosome

Chromosome arrangements





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Algorithm for computing Delaunay triangulation:

Input: N: number of points Ci: position of point I

1)Randomize points

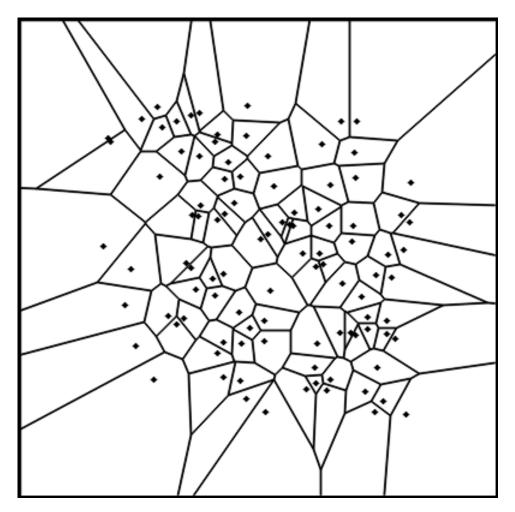
2) For i = 1:N

Location: find tetrehedra
that contains Ci

Addition: Divide t into 4

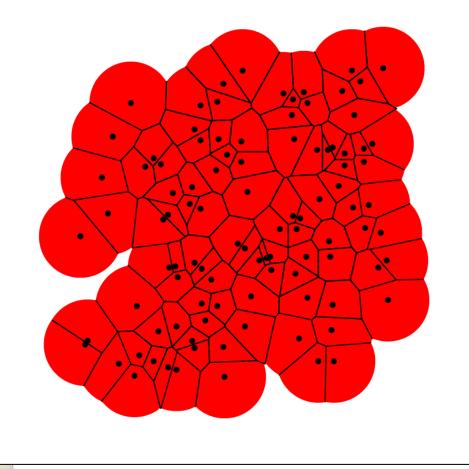
tetrahedra
Correct: flip non local tetrahedra

Output: list of tetrahedra



Compute Voronoi diagram from

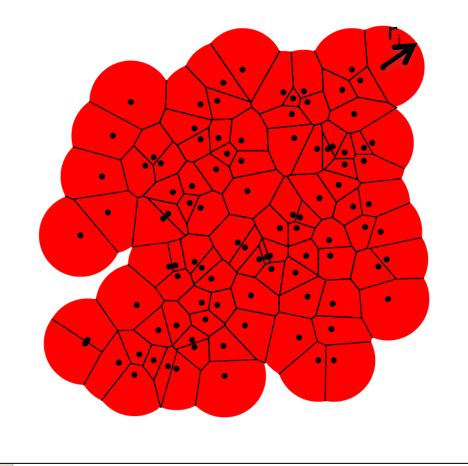
Delaaunay complex: dual



Restrict Voronoi diagram to

the Union of Balls:

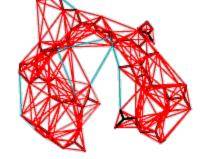
Power diagram



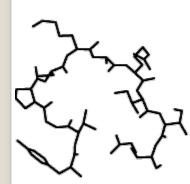
Atom i: Fraction in Voronoi cell: σ_i and β_i

 $A_i = 4\pi \sum_{i=1}^{N} r_i^2 \sigma_i$

 $V_i = \frac{4\pi}{3} \sum_{i=1}^N r_i^3 \beta_i$

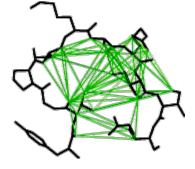


K complex

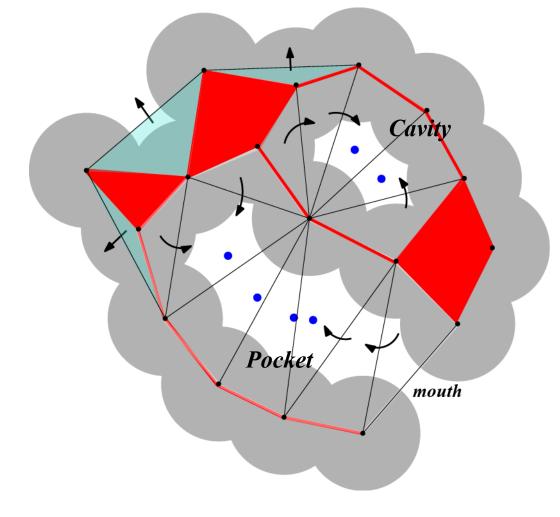


Protein

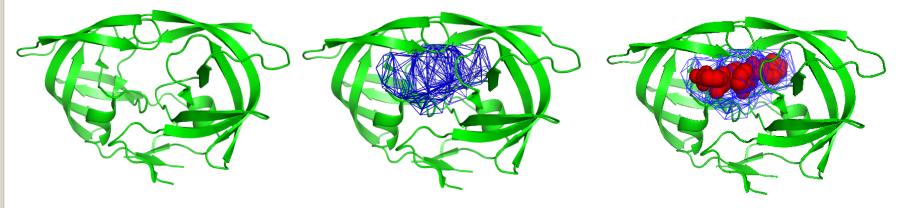
Delaunay Complex



Pocket



Applications to drug design

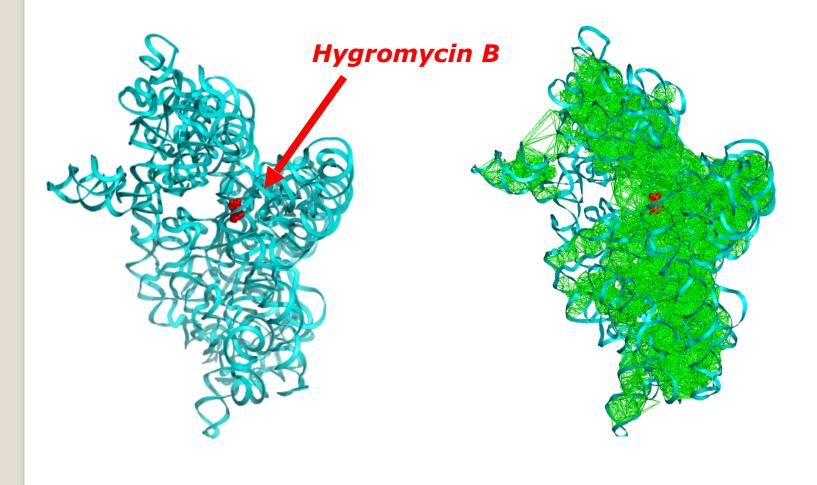


HIV protease (3MXE)

Main cavity

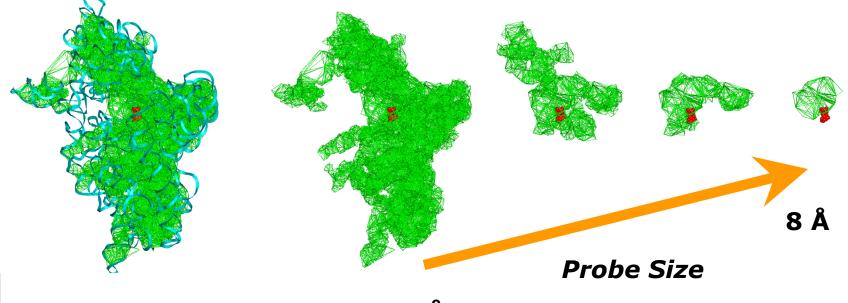
Actual position of K54 (inhibitor)

BINDING POCKETS IN 16S RIBOSOMAL RNA



PDB structure: 1HZN

BINDING POCKETS IN 16S RIBOSOMAL RNA



1.4 Å

Computing energy

Bonded interactions are local, and therefore their computation has a linear computational complexity (O(N), where N is the number of atoms in the molecule considered.

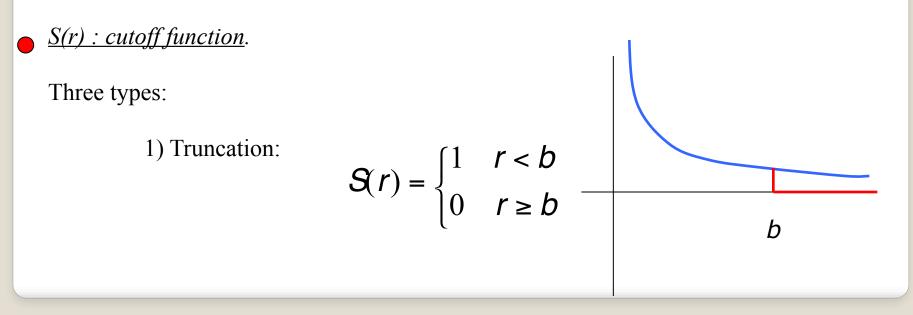
The direct computation of the non bonded interactions involve all pairs of atoms and has a quadratic complexity (O(N2)). This can be prohibitive for large molecules.

$$U_{NB} = \sum_{i, j \text{ nonbonded}} \varepsilon_{ij} \left[\left(\frac{R_{ij}}{r_{ij}} \right)^{12} - 2 \left(\frac{R_{ij}}{r_{ij}} \right)^{6} \right] + \sum_{i, j \text{ nonbonded}} \frac{q_i q_j}{4\pi\varepsilon_0 \varepsilon r_{ij}}$$

Cutoff schemes for faster energy computation

$$U_{NB} = \sum_{i,j} \omega_{ij} \mathcal{S}(\mathbf{r}_{ij}) \varepsilon_{ij} \left[\left(\frac{R_{ij}}{r_{ij}} \right)^{12} - 2 \left(\frac{R_{ij}}{r_{ij}} \right)^{6} \right] + \sum_{i,j} \omega_{ij} \mathcal{S}(\mathbf{r}_{ij}) \frac{q_{i}q_{j}}{4\pi\varepsilon_{0}\varepsilon r_{ij}}$$

 $\underline{\omega_{ij}: \text{weights}}_{or \ to \ scale \ some \ interactions} (usually \ 1-4)$



Cutoff schemes for faster energy computation

2. Switching

$$S(r) = \begin{cases} 1 & r < a \\ 1 + y(r)^2 [2y(r) - 3] & a \le r \le b \\ 0 & r > b \end{cases}$$

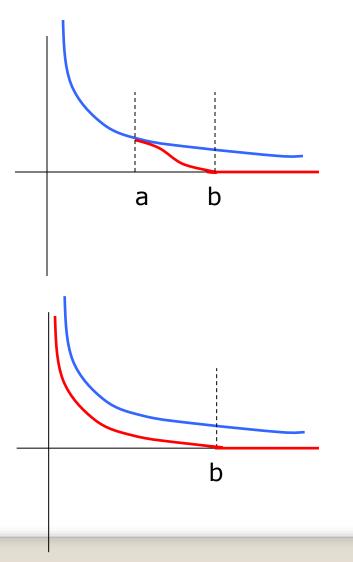
$$y(r) = \frac{r^2 - a^2}{b^2 - a^2}$$

3. Shifting

$$\mathbf{S}(\mathbf{r}) = \left[1 - \left(\frac{\mathbf{r}}{b}\right)^2\right]^2 \quad \mathbf{r} \le b$$

or

$$S_2(r) = \left[1 - \frac{r}{b}\right]^2 \quad r \le b$$



Units in Molecular Simulations

Most force fields use the AKMA (Angstrom – Kcal – Mol – Atomic Mass Unit) unit

system: Quantity AKMA unit Equivalent SI 1 Kcal/Mol Energy 4184 Joules Length 1 Angstrom 10⁻¹⁰ meter 1 amu 1.6605655 10⁻²⁷ Kg Mass (H=1amu) Charge 1 e 1.6021892 10⁻¹⁹ C 4.88882 10-14 Time 1 unit second 18.836 10¹⁰ rd/s Frequency 1 cm-1

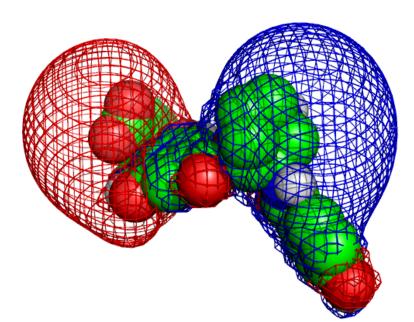
Some Common force fields in Computational Biology

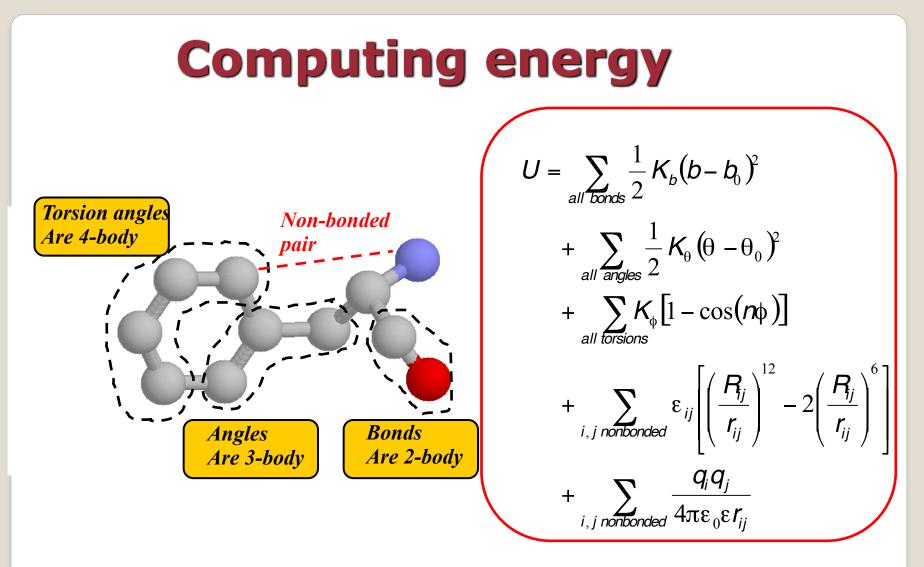
- ENCAD (Michael Levitt, Stanford)
- AMBER (Peter Kollman, UCSF; David Case, Scripps)
- CHARMM (Martin Karplus, Harvard)
- **OPLS** (Bill Jorgensen, Yale)
- MM2/MM3/MM4 (Norman Allinger, U. Georgia)
- ECEPP (Harold Scheraga, Cornell)
- **GROMOS** (Van Gunsteren, ETH, Zurich)

Michael Levitt. The birth of computational structural biology. Nature Structural Biology, 8, 392-(2001)

Biomolecular Simulations

- Molecular Mechanics force fields
- Energy Minimization
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U is a function of the conformation *C* of the protein. The problem of "minimizing *U*" can be stated as finding *C* such that *U*(*C*) is minimum.

The minimizers

Minimization of a multi-variable function is usually an iterative process, in which updates of the state variable x are computed using the gradient, and in some (favorable) cases the Hessian.

Iterations are stopped either when the maximum number of steps (user's input) is reached, or when the gradient norm is below a given threshold.

Steepest descent (SD):

The simplest iteration scheme consists of following the "steepest descent" direction:

$$\boldsymbol{X}_{k+1} = \boldsymbol{X}_{k} - \alpha \nabla f(\boldsymbol{X}_{k})$$

(α sets the minimum along the line defined by the gradient)

Usually, SD methods leads to improvement quickly, but then exhibit slow progress toward a solution.

They are commonly recommended for initial minimization iterations, when the starting function and gradient-norm values are very large.

The minimizers

Conjugate gradients (CG):

In each step of conjugate gradient methods, a search vector p_k is defined by a recursive formula:

$$p_{k+1} = -\nabla f(x_k) + \beta_{k+1} p_k$$

The corresponding new position is found by line minimization along p_k :

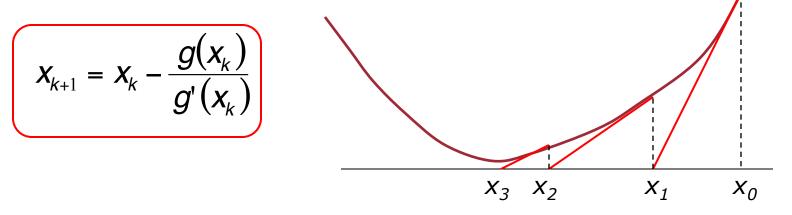
$$X_{k+1} = X_k + \lambda_k p_k$$

the CG methods differ in their definition of β .

The minimizers

Newton's methods:

Newton's method is a popular iterative method for finding the 0 of a one-dimensional function:



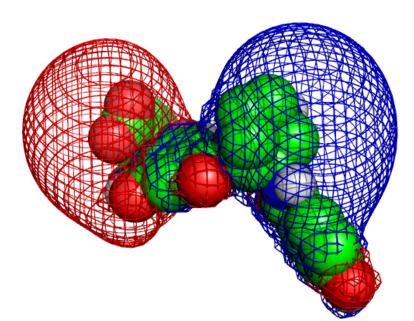
It can be adapted to the minimization of a one –dimensional function, in which case the iteration formula is: $x_{k} = x_{k} - \frac{g'(x_{k})}{g'(x_{k})}$

$$X_{k+1} = X_k - \frac{g'(x_k)}{g''(x_k)}$$

Several implementations of Newton's method exist: quasi-Newton, truncated Newton, "adopted-basis Newton-Raphson" (ABNR),...

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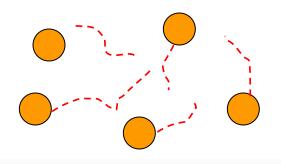


What is a molecular dynamics simulation?

Simulation that shows how the atoms in the system move with time

• Typically on the nanosecond timescale

 Atoms are treated like hard balls, and their motions are described by Newton's laws.



Characteristic protein motions

Type of motion	Timescale	Amplitude	_
Local:			
bond stretching	0.01 ps		$\llbracket \bigtriangleup \checkmark$
angle bending	0.1 ps	< 1 Å	
methyl rotation	1 ps		∐ V
Medium scale			' Periodic
loop motions			
SSE formation	ns – μs	1-5 Å	
Global			\sim
protein tumbling	20 ns	0	
(water	(20 ps)	> 5 Å	Random
tumbling) protein folding	ms – hrs	Y	
protein rolaing			

Why MD simulations?

- Link physics, chemistry and biology
- Model phenomena that cannot be observed experimentally
- Understand protein folding...
- Access to thermodynamics quantities (free energies, binding energies,...)

How do we run a MD simulation?

Get the initial configuration

From x-ray crystallography or NMR spectroscopy (PDB)

Assign initial velocities

At thermal equilibrium, the expected value of the kinetic energy of the system at temperature T is:

$$\langle E_{kin} \rangle = \frac{1}{2} \sum_{i=1}^{3N} m_i v_i^2 = \frac{1}{2} (3N) k_B T$$

This can be obtained by assigning the velocity components vi from a random Gaussian distribution with mean 0 and standard deviation (k_BT/m_i) :

$$\left\langle V_{i}^{2}\right\rangle = \frac{k_{B}T}{m_{i}}$$

How do we run a MD simulation?

• For each time step:

• Compute the force on each atom: $F(X) = -\nabla E(X) = -\frac{\partial E}{\partial X}$ X: cartesian vector of the system

 Solve Newton's 2nd law of motion for each atom, to get new coordinates and velocities

$$\overset{\bullet}{X} = F(X)$$

M diagonal mass matrix

.. means second order differentiation with respect to time

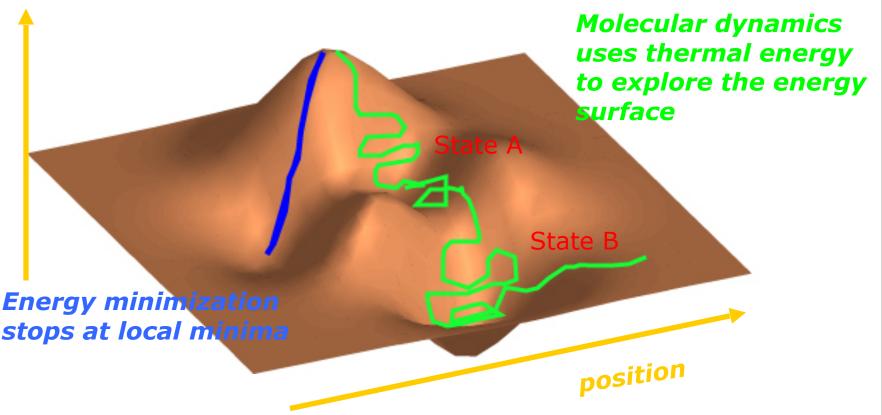
Store coordinates

Newton's equation cannot be solved analytically: —— Use stepwise numerical integration

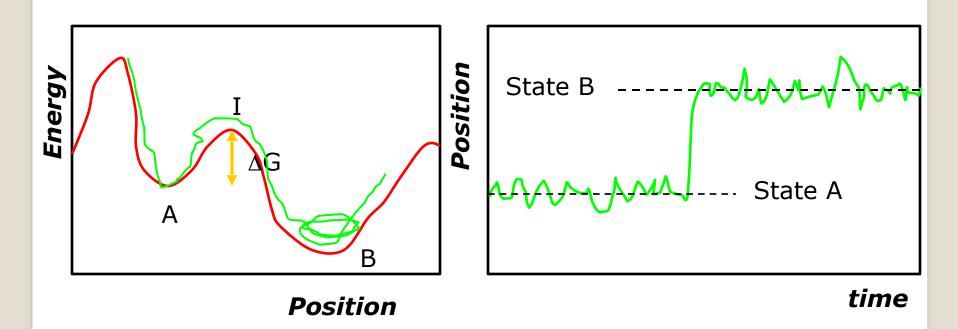
Stop

MD as a tool for minimization

Energy



Crossing energy barriers



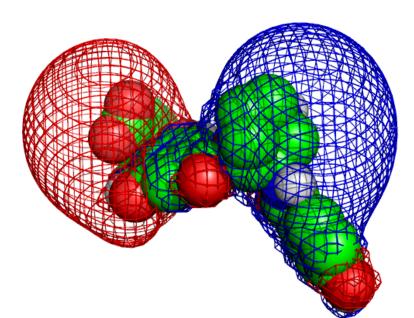
The actual transition time from A to B is very quick (a few pico seconds).

What takes time is waiting. The average waiting time for going from A to B can be expressed as: ΛG

$$\tau_{A \to B} = C e^{\frac{\Delta C}{kT}}$$

Biomolecular Simulations

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Monte Carlo: random sampling

A simple example:

Evaluate numerically the one-dimensional integral:

$$I = \int_{a}^{b} f(x) dx$$

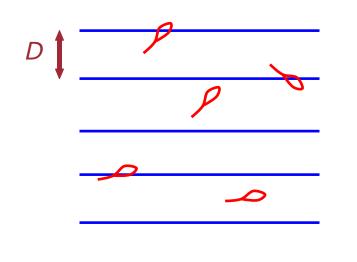
Instead of using classical quadrature, the integral can be rewritten as

$$I = (b-a)\langle f(x) \rangle$$

<f(x)> denotes the unweighted average of f(x) over [a,b], and can be determined by evaluating f(x) at a large number of x values randomly distributed over [a,b]

Monte Carlo method!

A famous example: Buffon's needle problem



The probability that a needle of length L overlaps with one of the lines, distant from each other by D, with $L \le D$ is:

$$P = \frac{2L}{\pi D}$$
For $L = D$

$$P = \frac{2}{\pi}$$

Method to estimate π numerically:

"Throw" N needles on the floor, find needles that cross one of the line (say C of them). An estimate of π is:

$$\pi = 2\frac{N}{C}$$

Buffon, G. Editor's note concerning a lecture given by Mr. Le Clerc de Buffon to the Royal Academy of Sciences in Paris. Histoire de l'Acad. Roy. des Sci., pp. 43-45, 1733. Buffon, G. "Essai d'arithmétique morale." Histoire naturelle, générale er particulière, Supplément 4, 46-123, 1777

Monte Carlo Sampling for protein structure

The probability of finding a protein (biomolecule) with a total energy E(X) is:

$$P(X) = \int \exp\left(-\frac{E(X)}{kT}\right) dZ \longrightarrow Partition function$$

Estimates of any average quantity of the form:

$$\langle A \rangle = \int A(X) P(X) dX$$

using uniform sampling would therefore be extremely inefficient.

Metropolis and coll. developed a method for directly sampling according to the actual distribution.

Metropolis et al. Equation of state calculations by fast computing machines. J. Chem. Phys. 21:1087-1092 (1953)

Monte Carlo for sampling conformations

The Metropolis Monte Carlo algorithm:

- 1. Select a conformation X at random. Compute its energy E(X)
- 2. Generate a new trial conformation Y. Compute its energy E(Y)
- 3. Accept the move from X to Y with probability:

$$P = \min(1, \exp\left(-\frac{E_{\rho}(Y) - E_{\rho}(X)}{kT}\right)$$

Pick a random number RN, uniform in [0,1]. *If RN < P, accept the move.*

4. Repeat 2 and 3.